

Using statistical profiling to decipher hidden chromatin contacts resulting from repeated sequences

26/11/2024

Sébastien Gradit

Spatial Regulation of Genomes - Genomes & Genetics - UMR3525

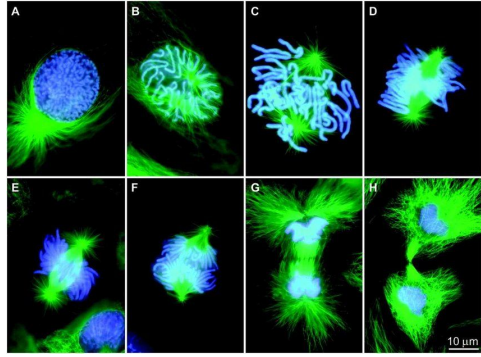
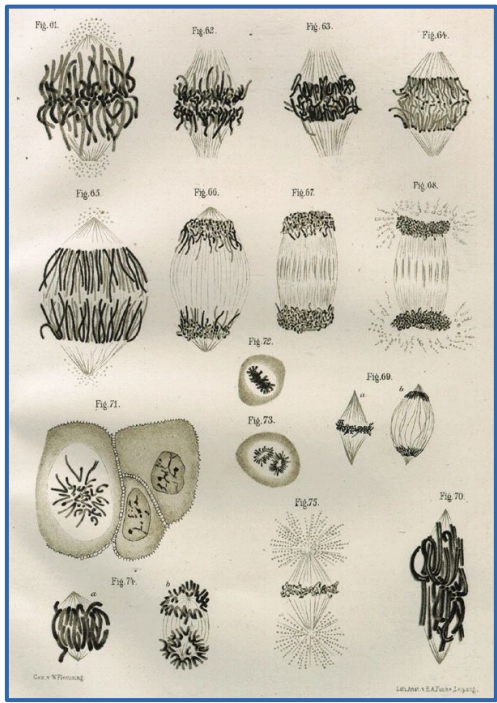
Thesis supervisor: Axel Cournac



Early observation of genomes organisation



Lilium corceum
drawings



DNA folding is a highly organized process necessary to fit DNA in nucleus

Human cell



10 µm

$3 \cdot 10^9$ bp = 2m

yeast

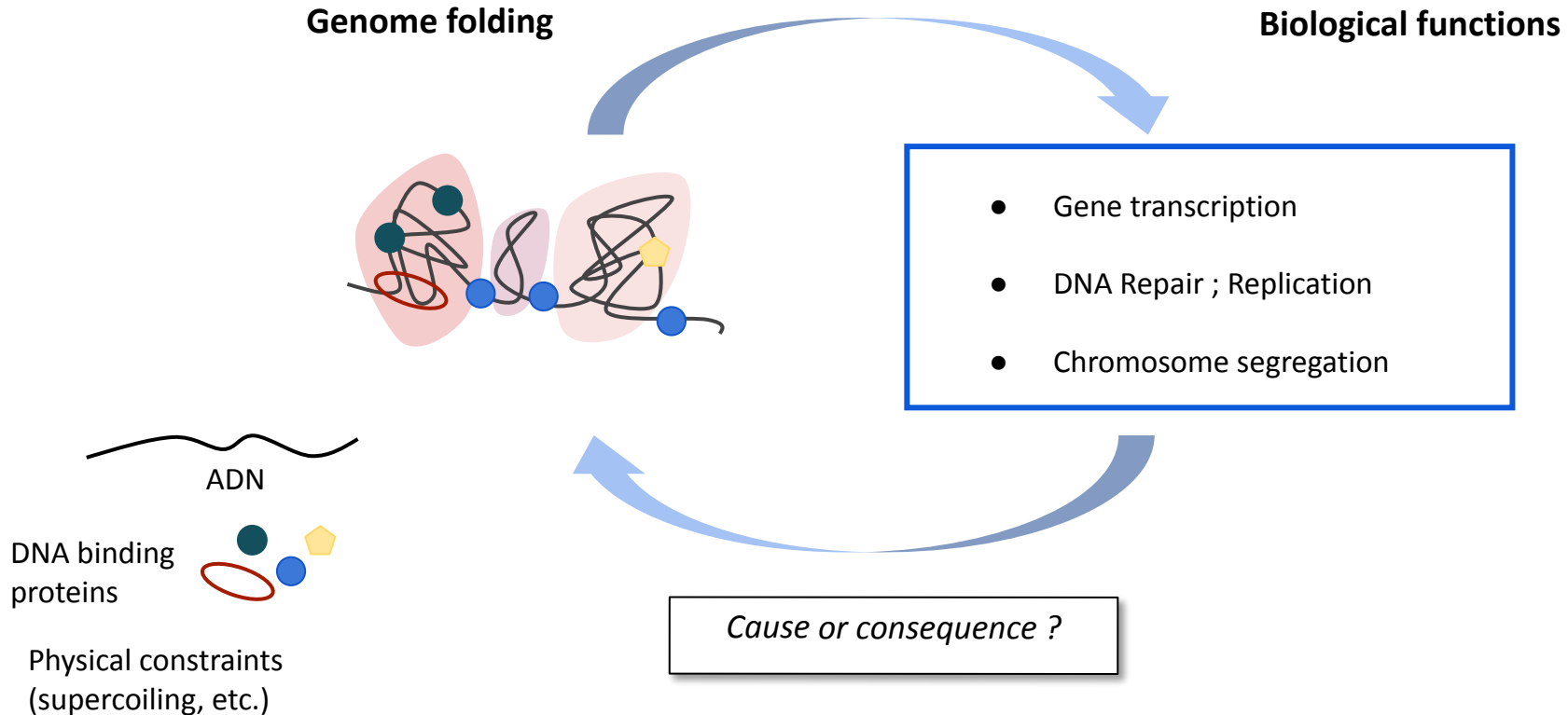


2 µm

$12 \cdot 10^6$ bp = 8 mm

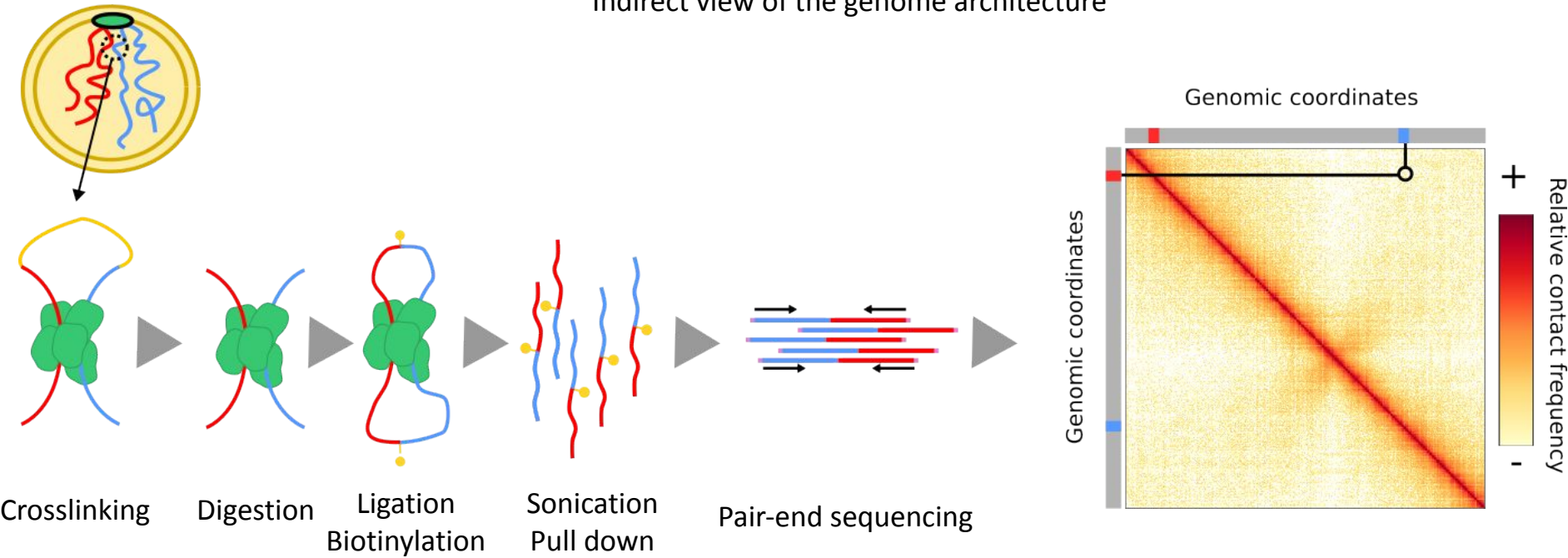
Zellsubstanz, Kern und Zelltheilung, Flemming, 1882
Risler, 2011

Chromosome folding influences or regulates dynamic processes?



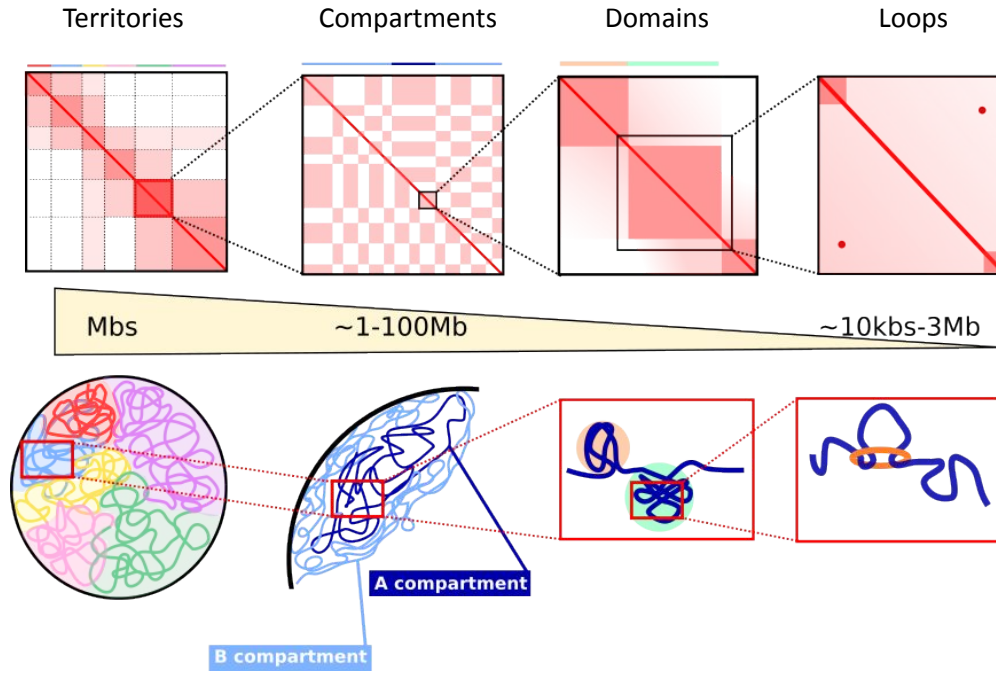
Hi-C to capture genomes 3D structures

Indirect view of the genome architecture



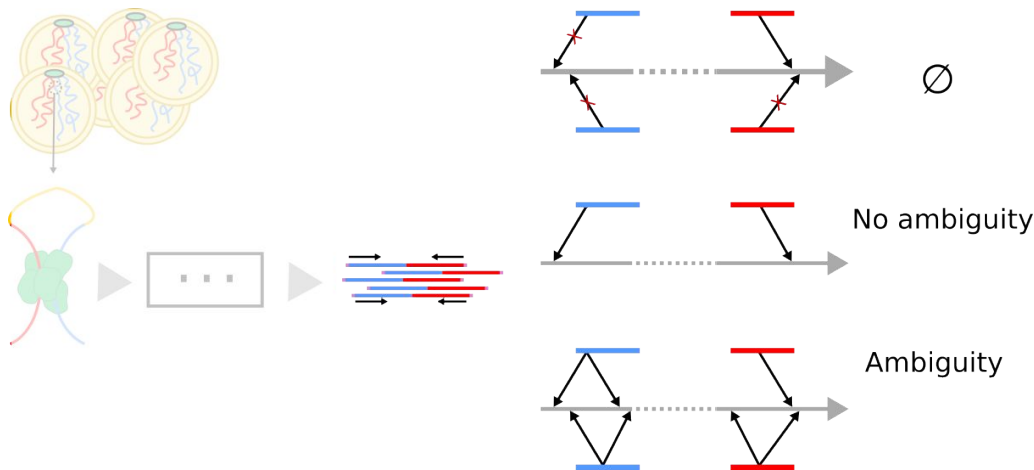
Chromosome Configurations: Influencing Genome Organization

How these different spatial structures correlates with biological processes?

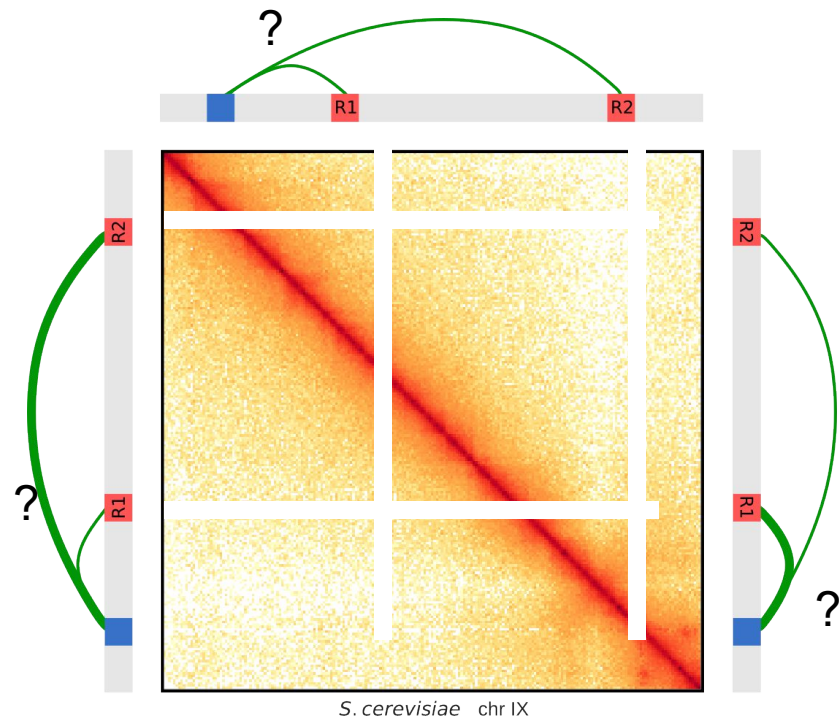


- Cell cycle phase dependence
- Response to stresses
- Quiescence / replicating?
- Viral infection?

Limitations of Traditional Hi-C

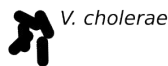
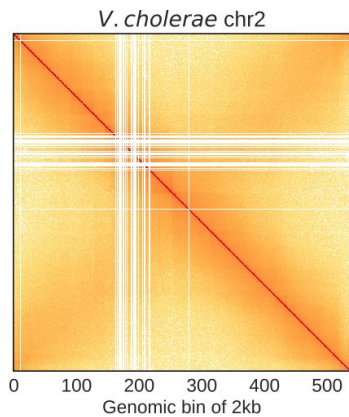


- Pair-end sequencing produces short reads ~ 50 bp
- Genomes carry repeated regions
→ Some reads can have multiple genomic positions

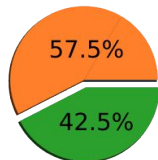


Statistical approach for ambiguous contacts prediction?

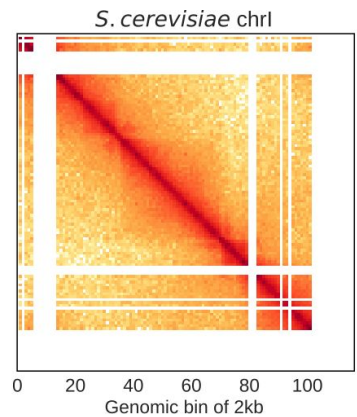
Thesis Objectives



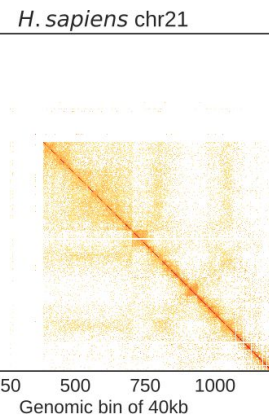
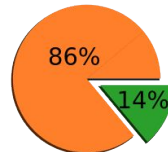
V. cholerae



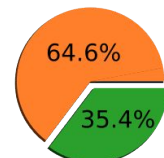
Non repeated sequences
Repeated sequences



S. cerevisiae



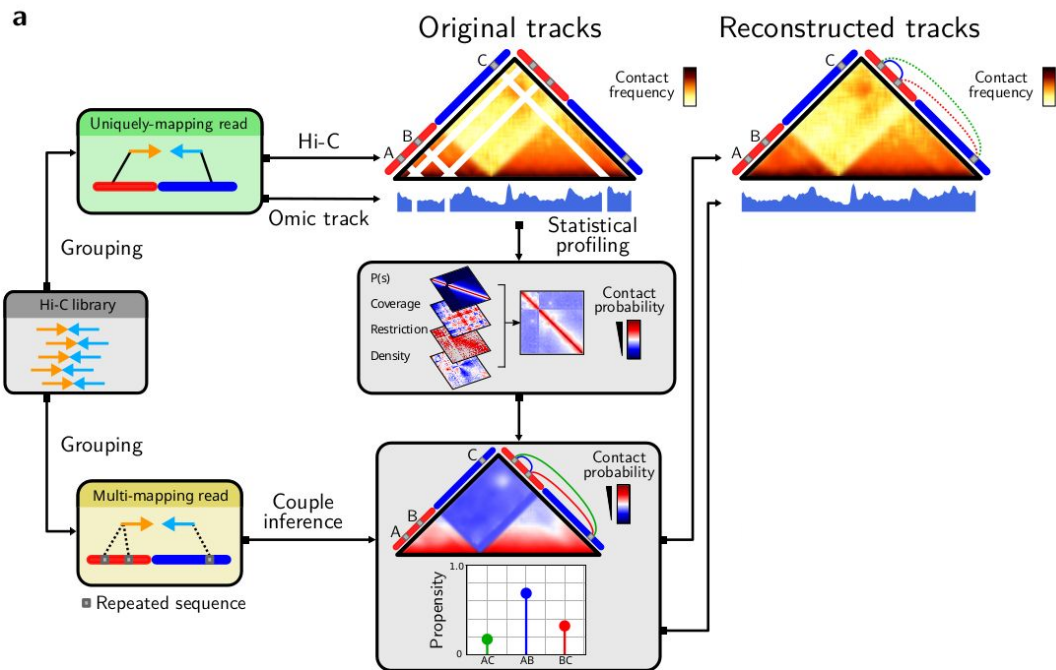
H. sapiens



Can a statistical modeling approach be used to infer missing information in Hi-C data and accurately reconstruct the 3D interactions of repeated sequences?

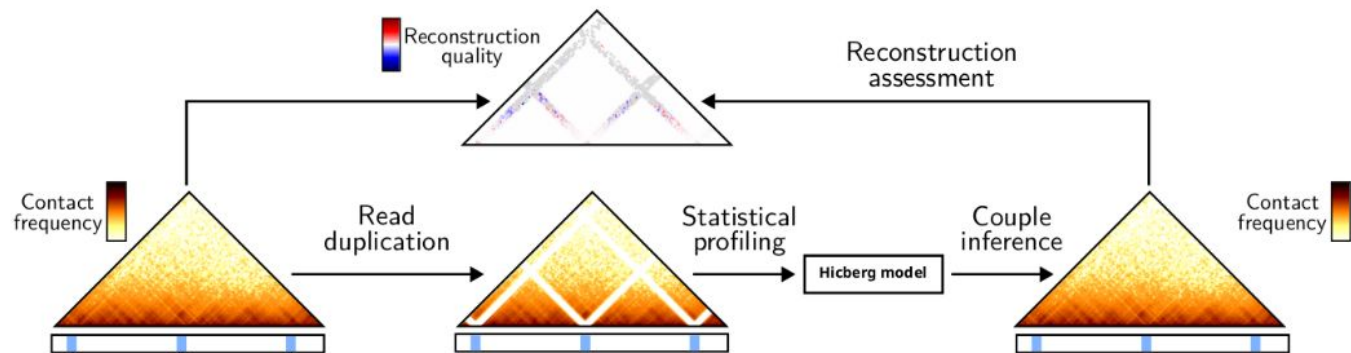
Hicberg: Reconstruction of genomic signals from repeated elements

Hicberg: a novel tool for (3D) genomics



- **Hicberg:** Reconstruction of Hi-C data, and more!
- **On-the-fly statistical profiling:** extraction of statistical tendencies from unambiguous data.
- **Probability Mass Function estimation:** Guides accurate placement of ambiguous reads.

Benchmarking & In-silico Evaluation

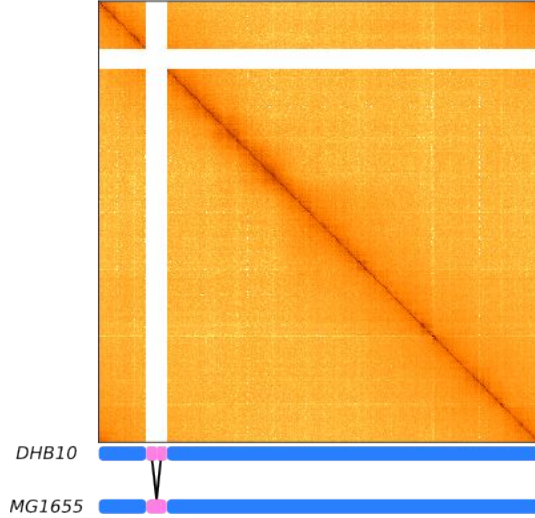


- **Simulated** repeated sequences by duplicating unambiguous reads.
- **Robust** across various repeat sizes, numbers, and spacing.
- **Polymeric** behavior component is key for accurate inference.

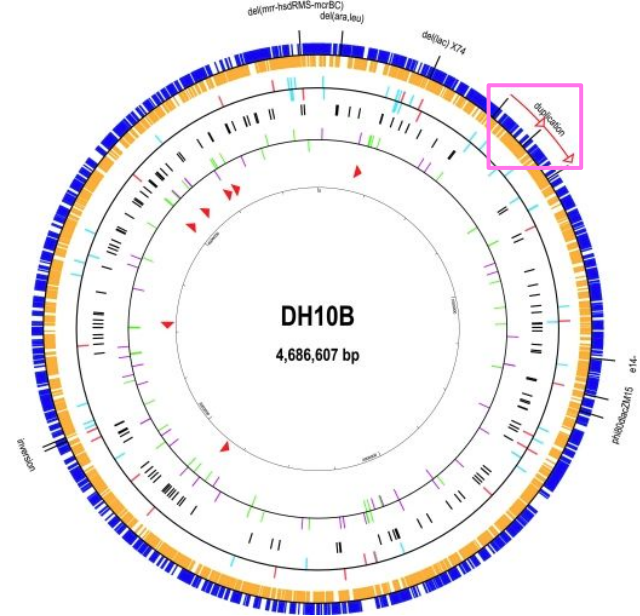
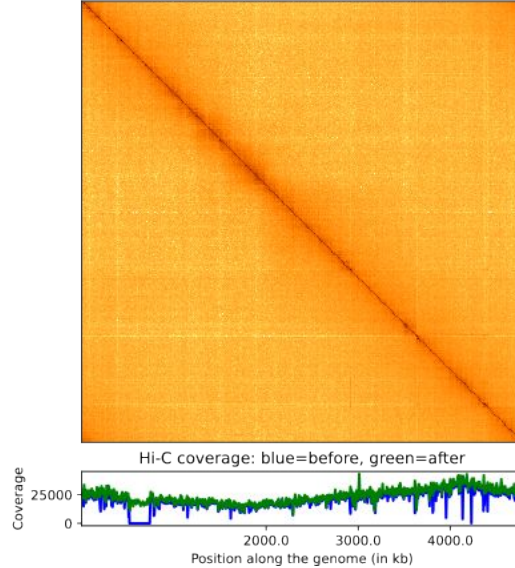
Does Hicberg perform as well in real-context?

Biological validation

Contacts map CalTech4

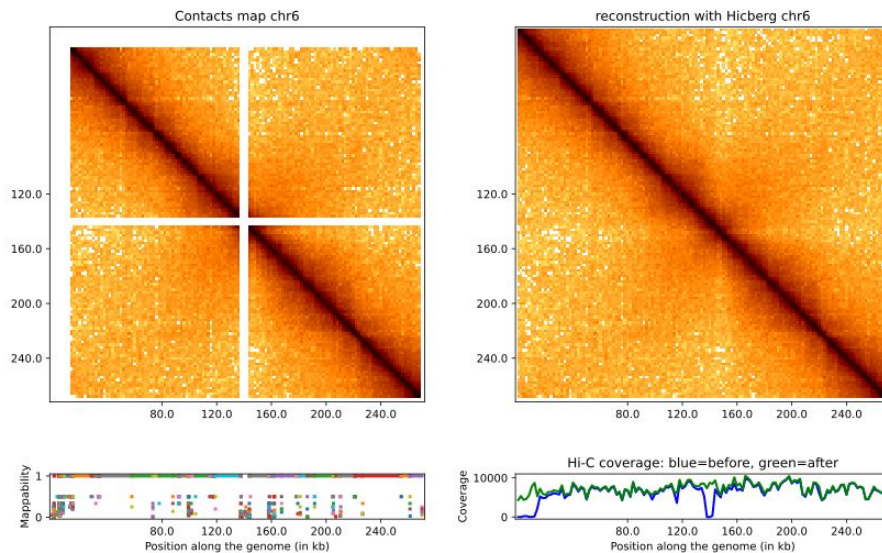


reconstruction with Hicberg CalTech4



- Accurate reconstruction of the complex 3D structure of *E. coli* carrying tandem duplication.
- Hicberg captured expected DNA polymer behavior, demonstrating versatility and power.

Biological validation



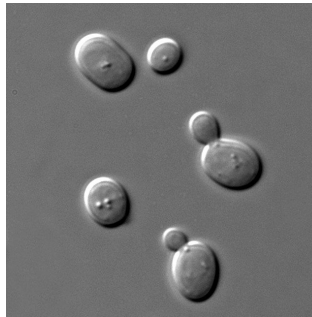
- Accurate reconstruction of the complex 3D structure of *S. cerevisiae* chr VI carrying repeats.
- Hicberg captured expected DNA polymer behavior, demonstrating versatility and power.
- Hicberg recovered expected mean coverage despite low theoretical mappability

Unveiling Hidden Connections: Hicberg in action

How do repetitive elements shape the 3D genome and influence its function?

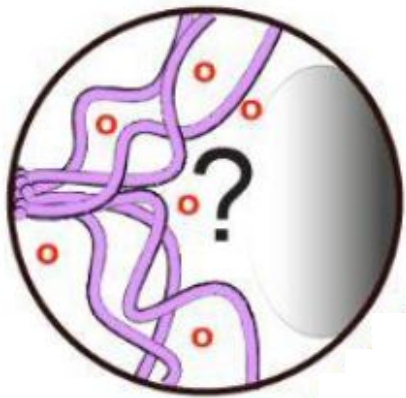
1) *Study of Ty1 Retrotransposons: 3D Organization and Cohesin Loading*

2) *Study of rDNA Contact Dynamics in Response to Stress*

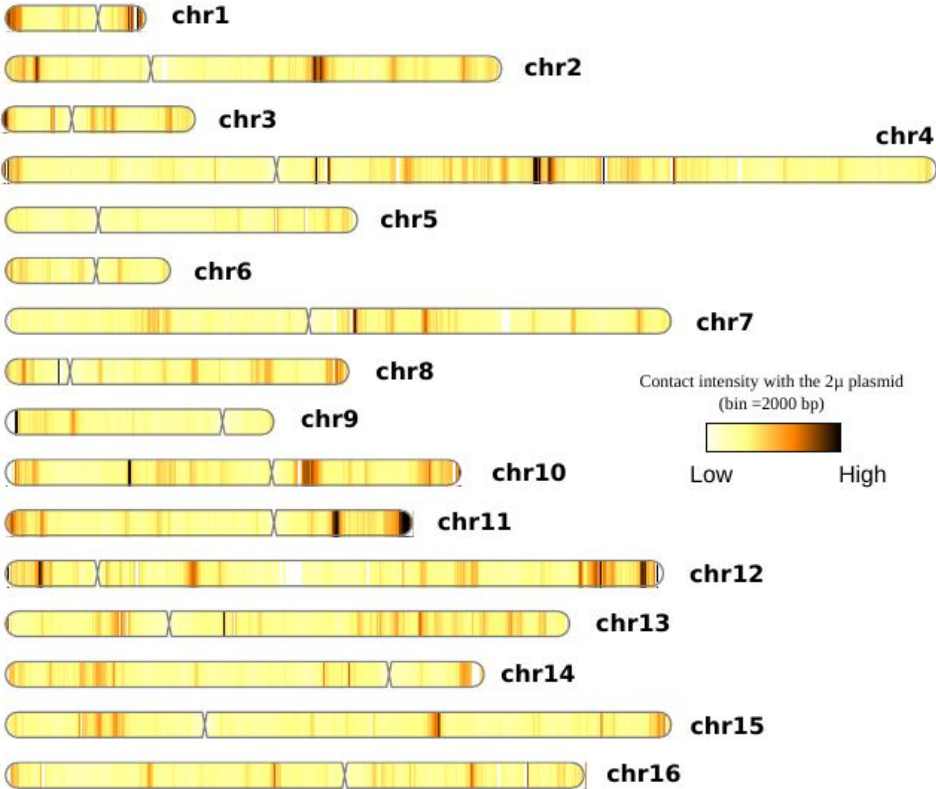


Beyond Parasitic Elements: Ty1 Retrotransposons as Organizers of Nuclear Architecture

Application : *S. cerevisiae* TYs as new 2μ preferential contact spots

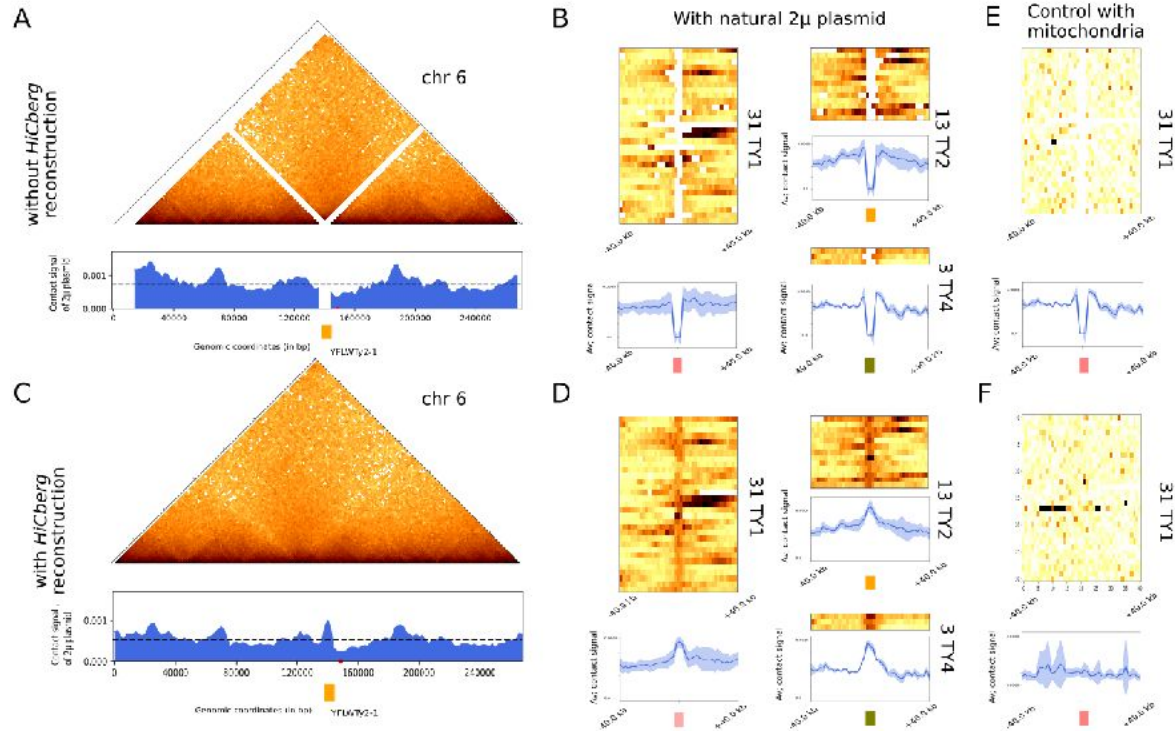


Presence of natural 2μ plasmid in *S. cerevisiae*



Detection of ~ 75 hotspots of contact

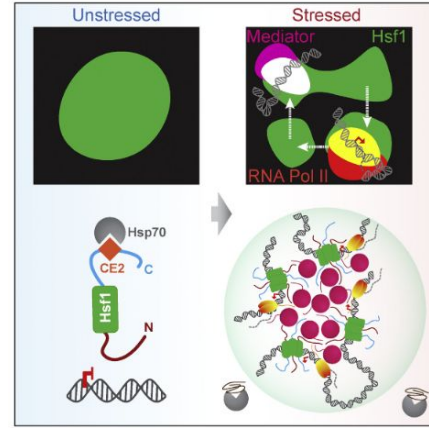
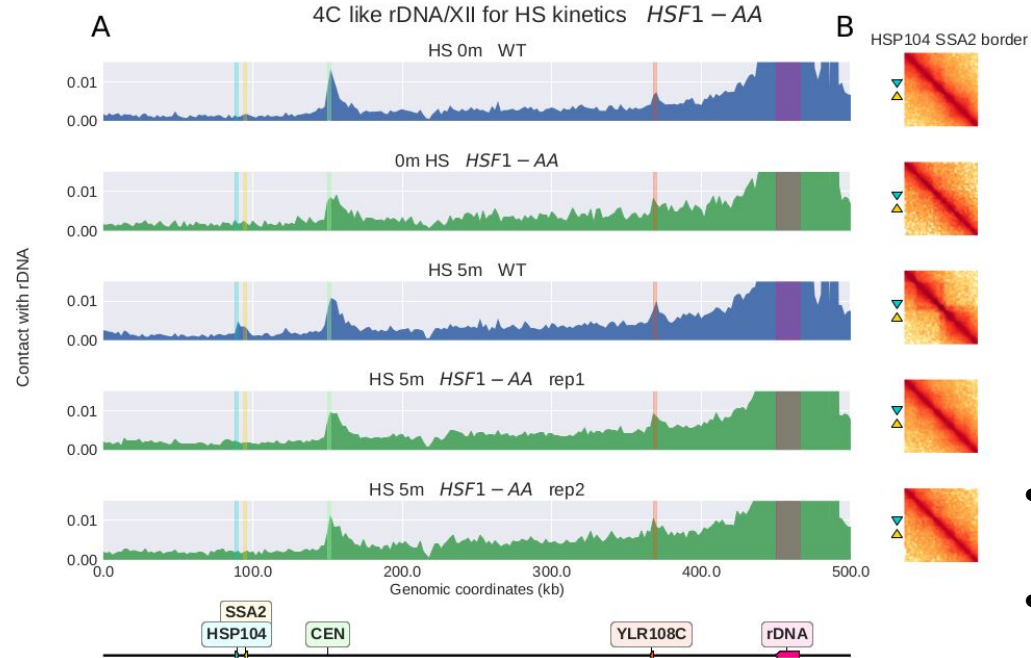
Application : *S. cerevisiae* TYs as new 2 μ preferential contact spots



Retrotransposons (TYs) appear to be new hotspots of 2 μ plasmid

Unveiling the rDNA Network: Key Players and Choreographers of its Spatial Organization

Application : *S. cerevisiae* TYs as new 2 μ preferential contact spots



- **Hsf1 Mutant:** Transient rDNA-HSP104/SSA2 interaction and border pattern are lost.
- **Dual Requirement:** Both transcription and Hsf1 are essential for this stress-induced structure.
- **Mechanism:** Supports Chowdary's hypothesis of micro-aggregates, not loop extrusion, for contact formation.

- Hicberg showed **good performances** both in **simulated** and in **biological** data
- Several genomic features have been unveiled in *S. cerevisiae*:
 - 2μ plasmid **hotspots of contacts** of yeast retrotransposons
 - **New rDNA** interaction during **heat shock**, involving *Hsf1* protein complex

→ First link about spatial organization of rDNA and **stress**



Régulation Spatiale des Génomes

Romain Koszul
Axel Cournac
 Martial Marbouty
 Agnès Thierry
 Hélène Bordelet
 Jacques Serizay
Pauline Larrous
 Devon Conti
 Justine Groseille
 Manon Perrot
 Maëlys Delouis
 Corina Pascal

Anciens membres

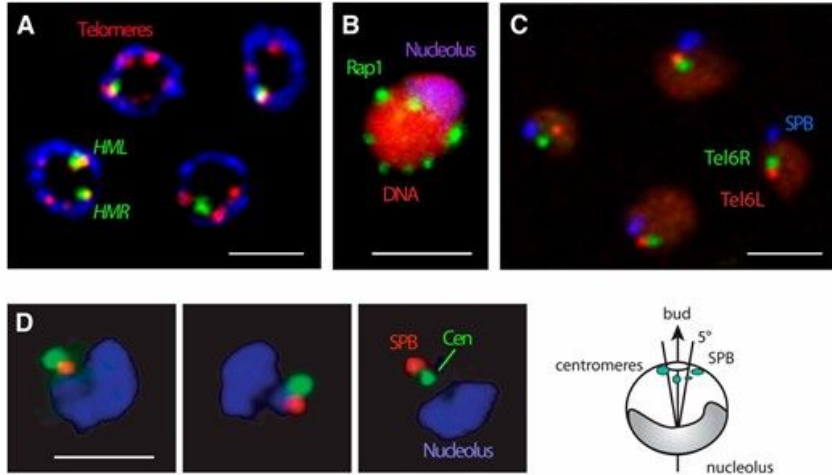
Cyril Matthey-Doret
 Amaury Bignaud
 Léa Meneu
 Samuel Ortion

Collaborators

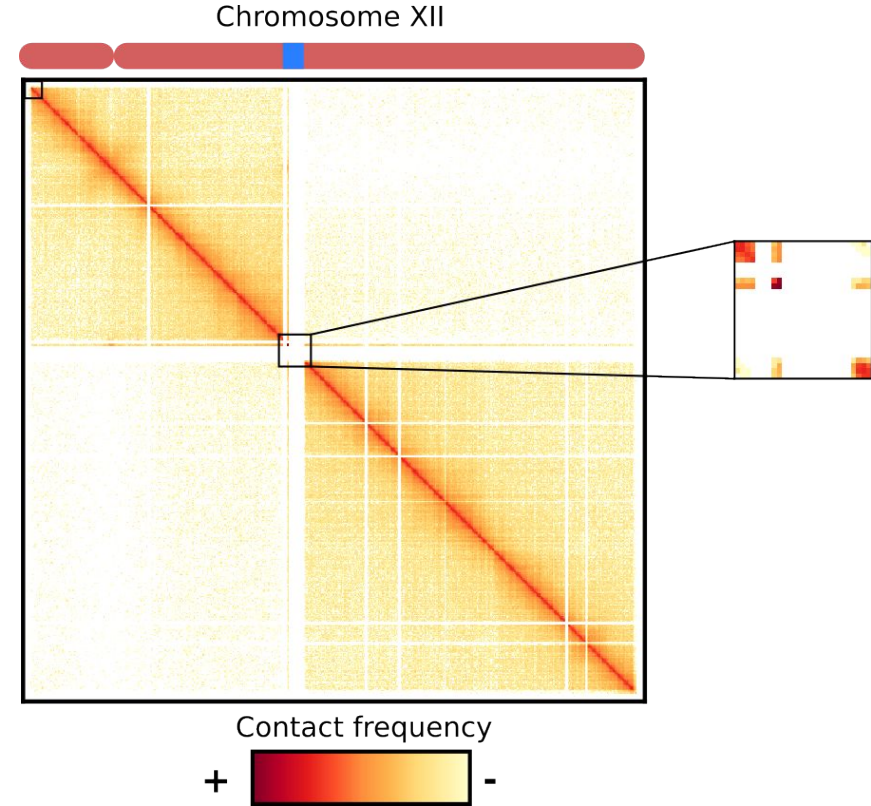
Bouk Wim De Jong



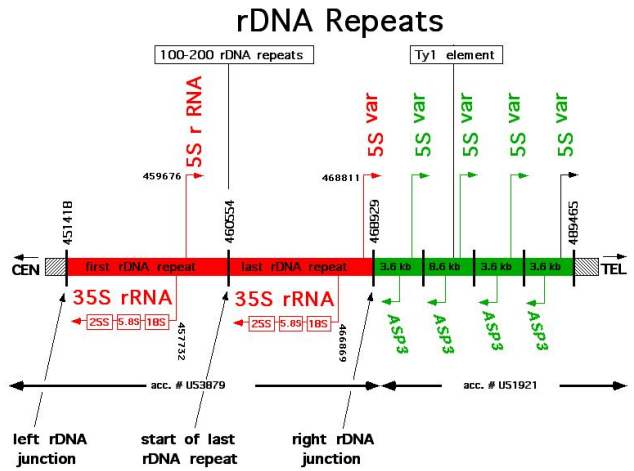
rDNA: A Landmark in the Nuclear Landscape



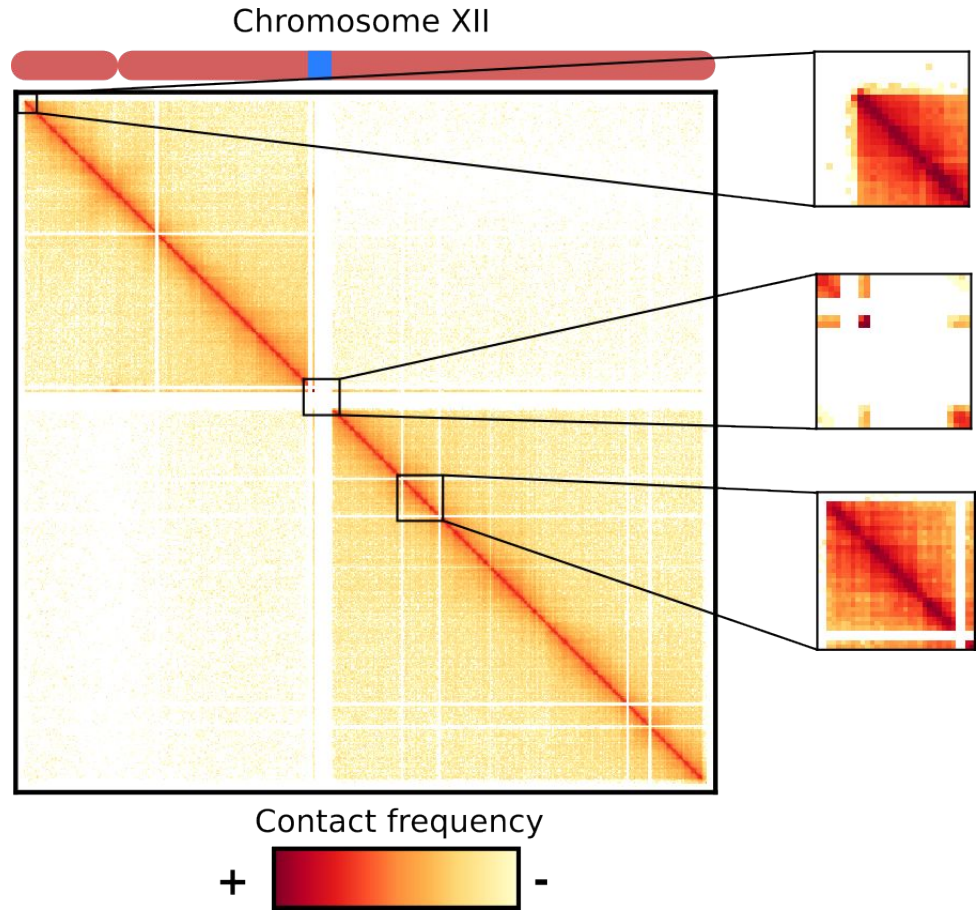
- rDNA as an organizational feature within the nucleus
- Ribosomes biogenesis
- Often exhibits interactions with nuclear landmarks (telomere, centromeres)



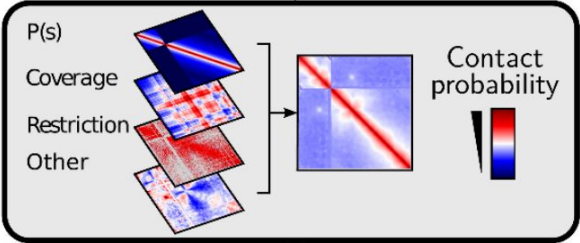
The Challenge of Repeated Sequences



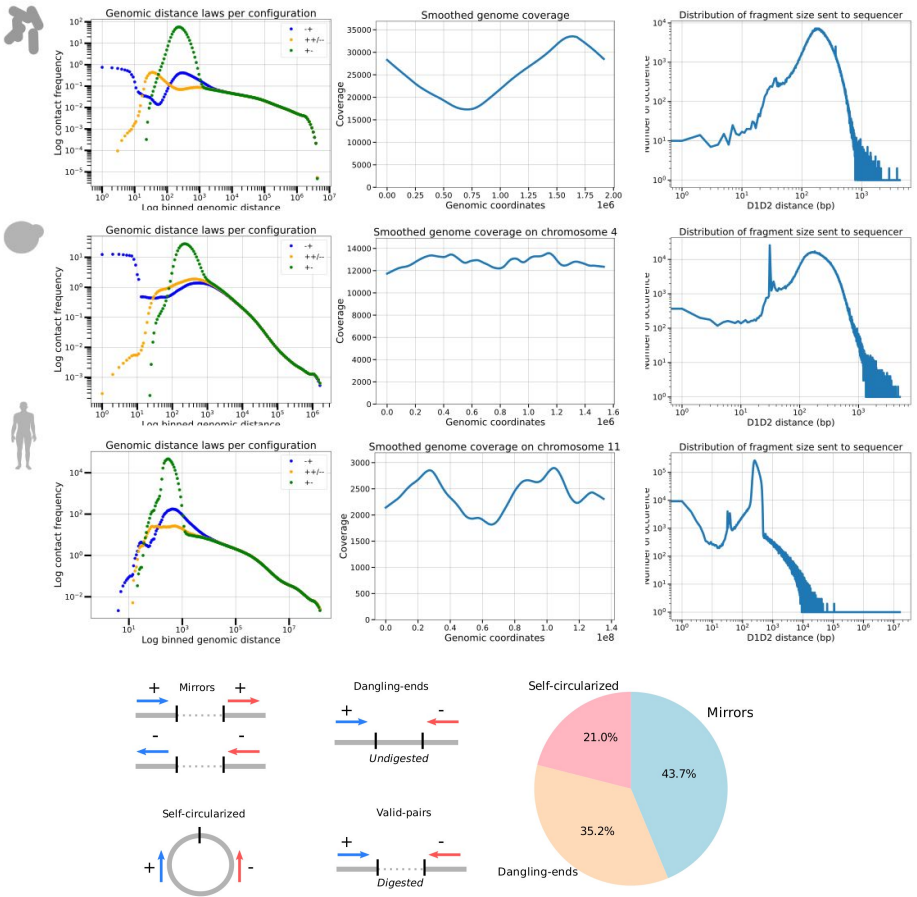
Why are repetitive sequences, particularly challenging to study using traditional Hi-C?



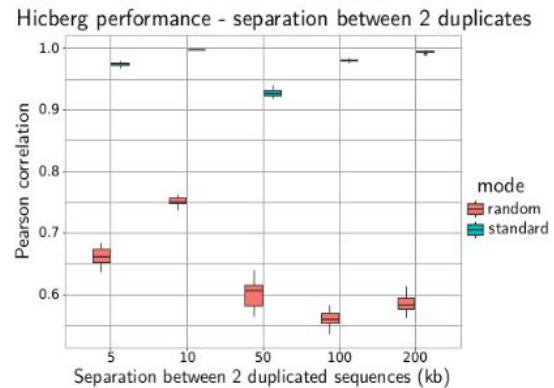
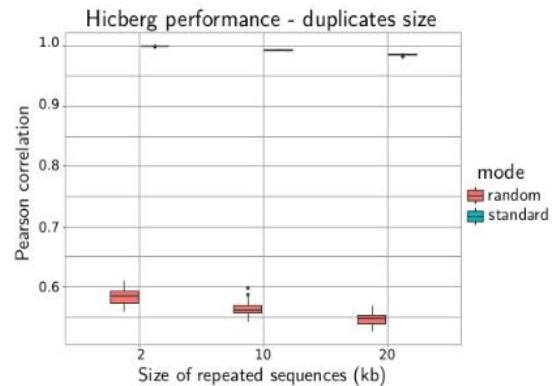
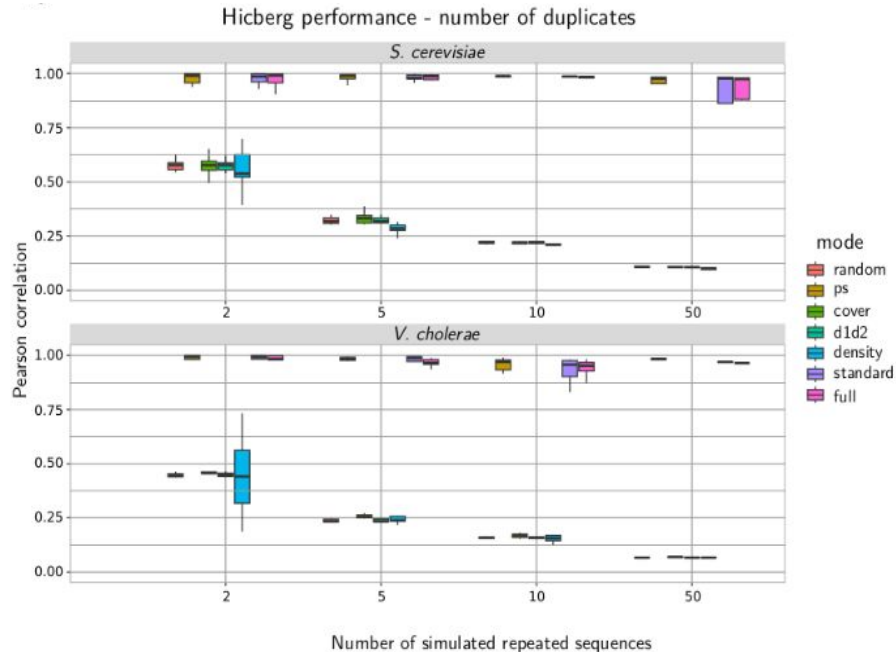
Hicberg: a novel tool for (3D) genomics



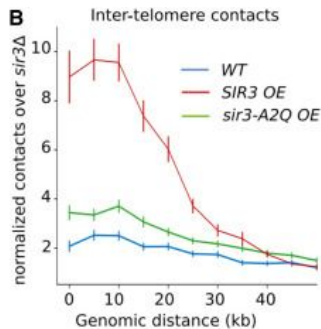
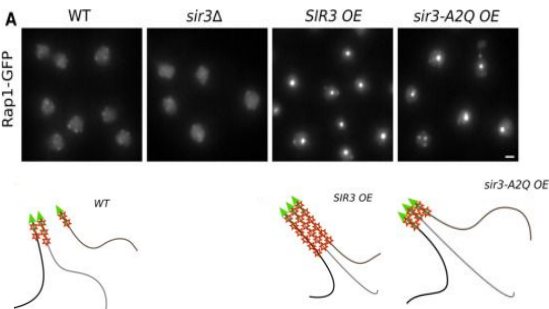
- Polymer Behavior:** Captures DNA flexibility with 3 species/condition-specific sub-laws.
- Coverage Bias:** Accounts for uneven read distribution; higher coverage increases interaction probability.
- Restriction Site Proximity:** Accounts for fragment length biases.



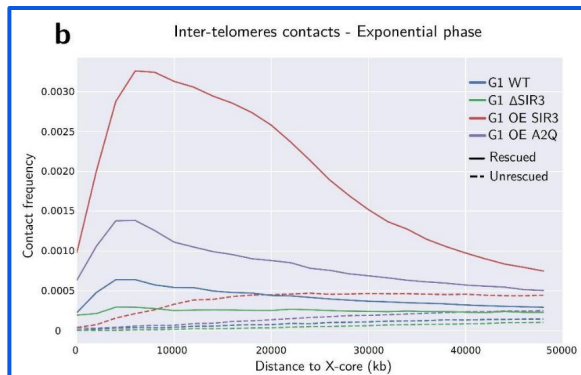
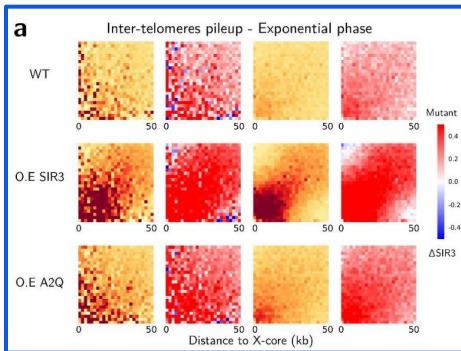
In-silico Evaluation



Biological validation



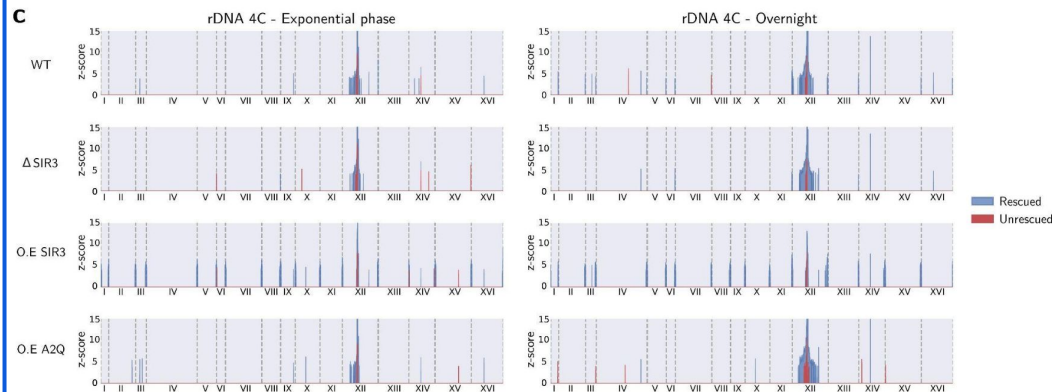
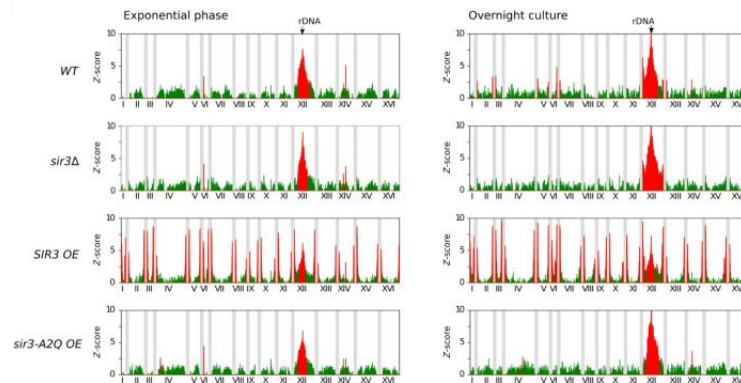
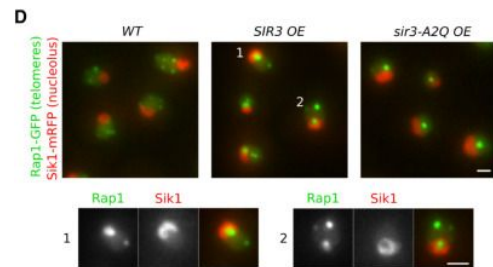
Hicberg to re-analyze yeast Hi-C data on telomere clustering under SIR3 over-expression studied with a diffusion-based method (Serpentine)



Hicberg reconstruction: Accurately reproduced Ruault's findings:

- SIR3 overexpression → telomere hyper-clustering.
- Altered SIR3 (A2Q) → reduced clustering.

Biological validation

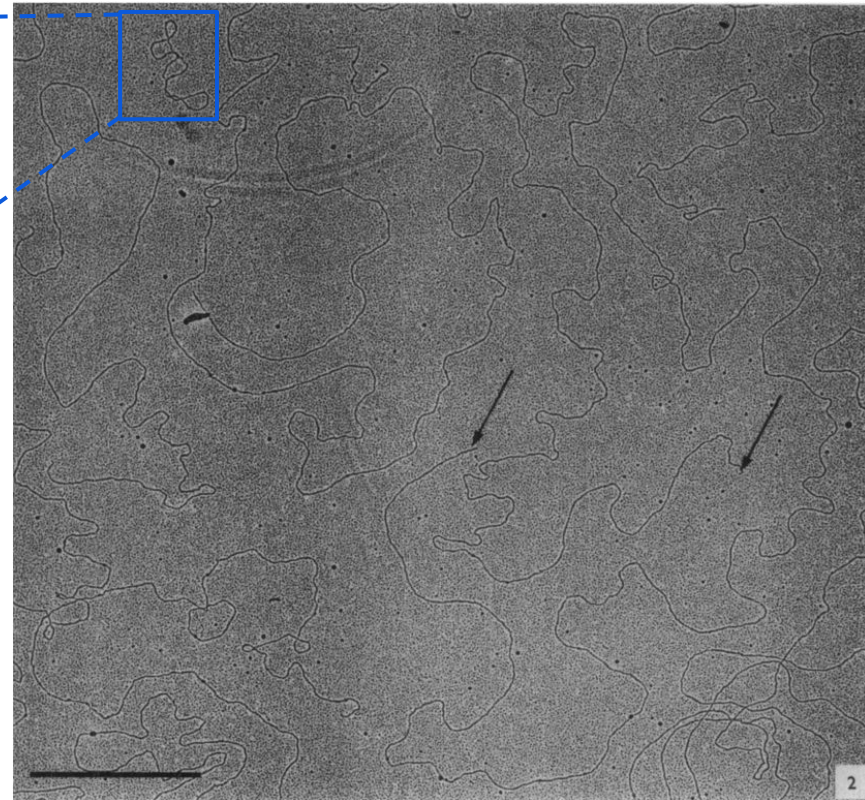
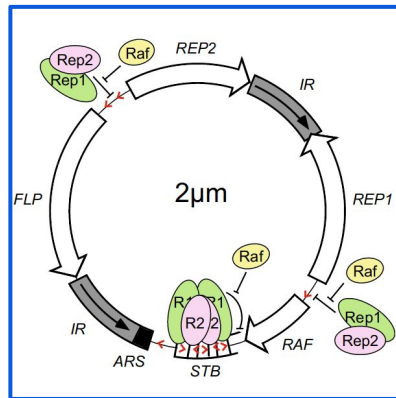


Hicberg reconstruction: Reproduced Ruault's findings:

- SIR3 overexpression → increased rDNA-telomere clustering (exponential and overnight cultures).
- Altered SIR3 (A2Q) → partially reduced clustering.

Hicberg accurately captures complex, condition-specific 3D interactions in repeat rich regions, further validating its biological relevance.

Discovery of 2μ plasmid in *Saccharomyces cerevisiae*



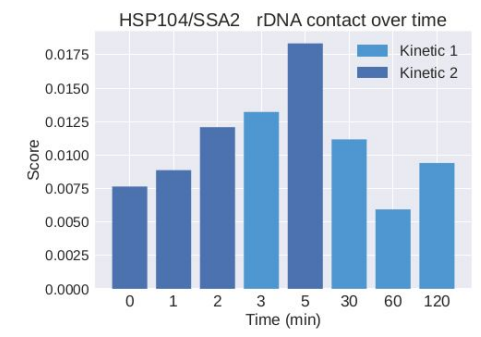
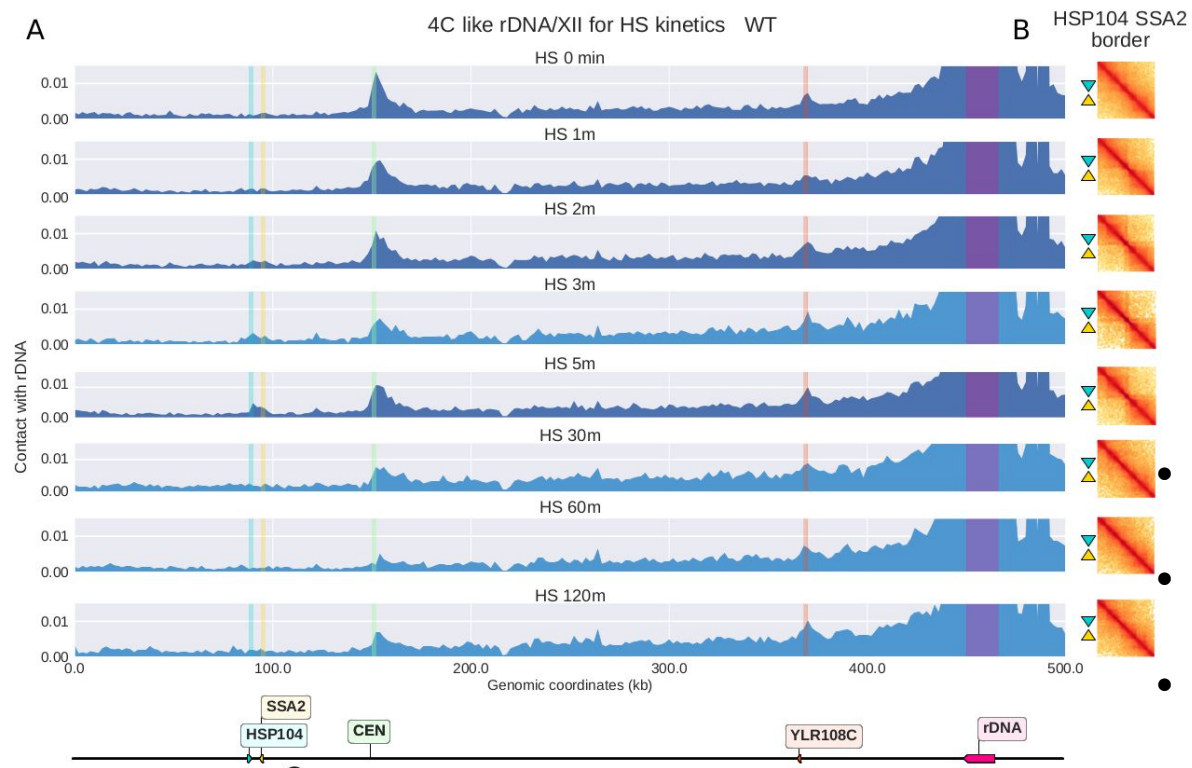
Observation in the 1970's of **enriched fraction of small circular DNA** found as different molecules species

Predominant class of with **mean contour length of $1.88 \pm 0.11 \mu\text{m}$**

Conserved through generations

Specific locations in *S.cerevisiae* genome (long genes, low transcription spots)

rDNA-HSP104/SSA2: A Transient Tango During Heat Shock



- **rDNA in Heat Shock:** Hicberg reveals transient interaction with HSP104/SSA2 genes.
- **Dynamic contact:** Forms and dissolves during heat shock, suggesting a role in stress response.
- **Border pattern:** Emerges around HSP104/SSA2, indicating potential transcriptional activation.

HiC-BERG - Conclusion & Perspectives

- Hicberg showed good performances both in simulated and in context data
 - Hicberg outperformed mHiC on the definition of reconstructed structures, and number of retrieved Hi-C reads
 - Several genomic features have been unveiled (2 μ hotspots on TYs, new rDNA interaction during heat shock, transcriptionally induced aggregates)
 - First link about spatial organization of rDNA and stress
-

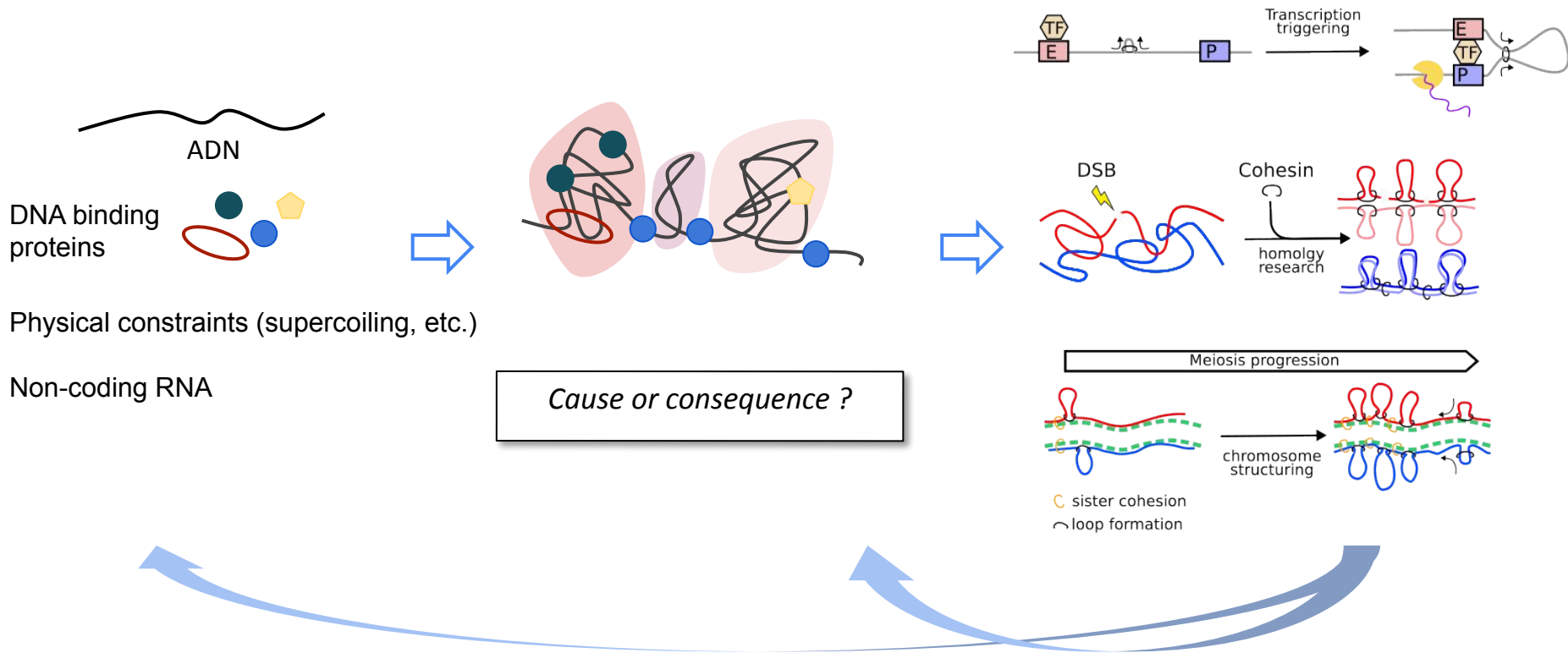
Expand Applications:

- Explore diverse stress conditions in yeast.
- Apply to other organisms and datasets.
- Investigate various "OMICS" data.

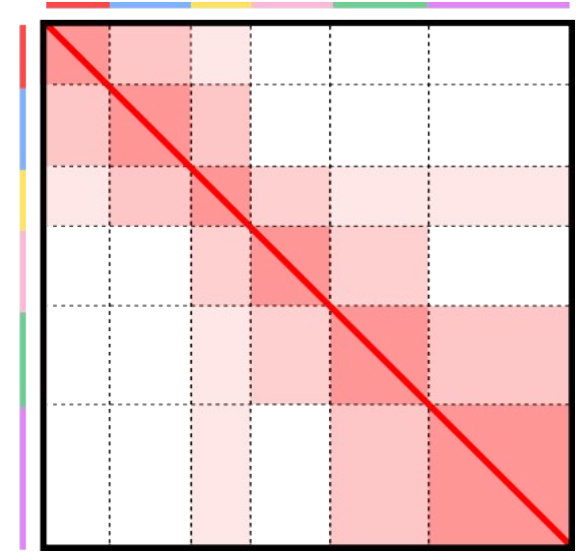
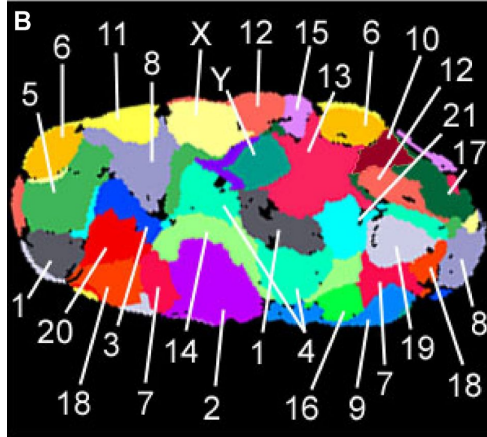
Enhance Performance:

- Optimize for speed and efficiency.
- Facilitate routine use across species.

Chromosome folding influences or regulates dynamic processes?



Chromosome Location in Genomes

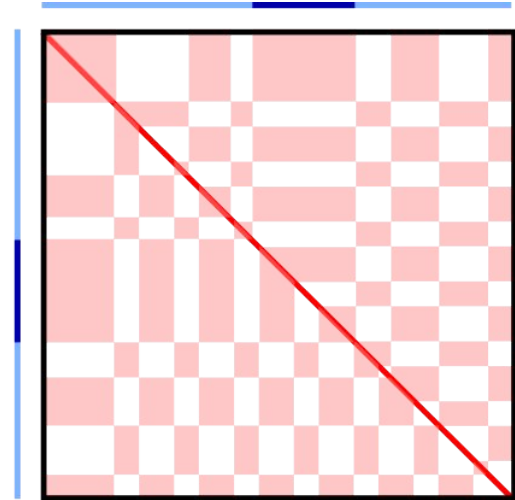
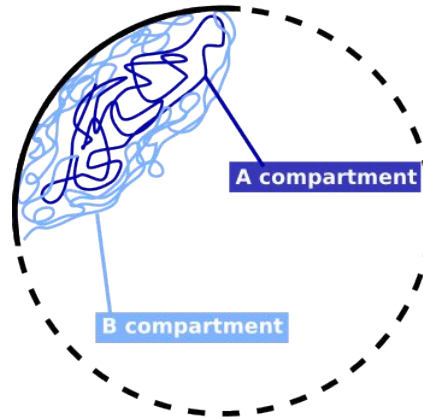
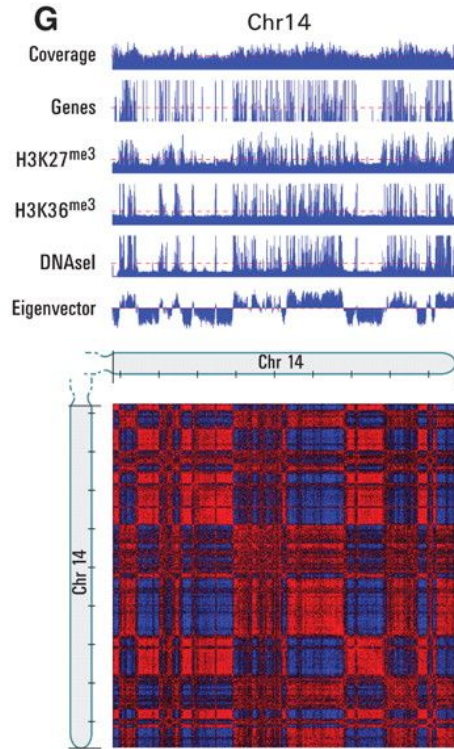


- Size/shape vary → chromosome, cell type
- Dynamic state (cellular processes, cycle, ...)
- Gene rich toward interior

Response to biological processes:

- Efficiency of DNA replication and repair
- Prevent inter-chromosomes translocations

From chromosomes to compartments



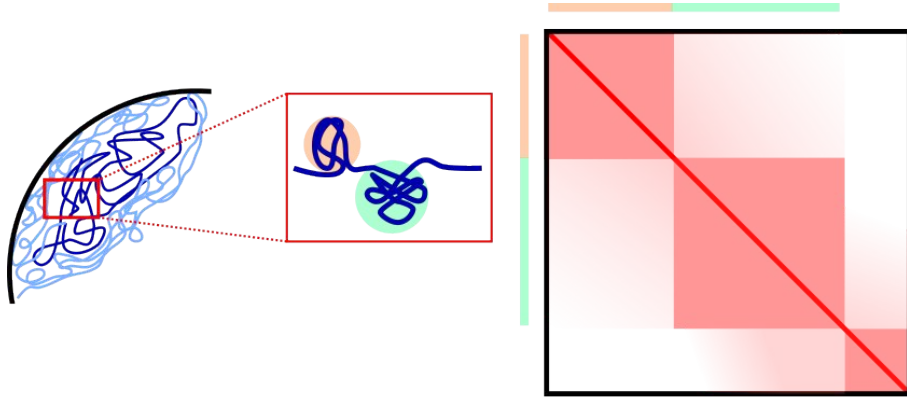
A compartments:

- Open chromatin
- Gene rich
- Acetylation marks
- Interior of nucleus

B compartments:

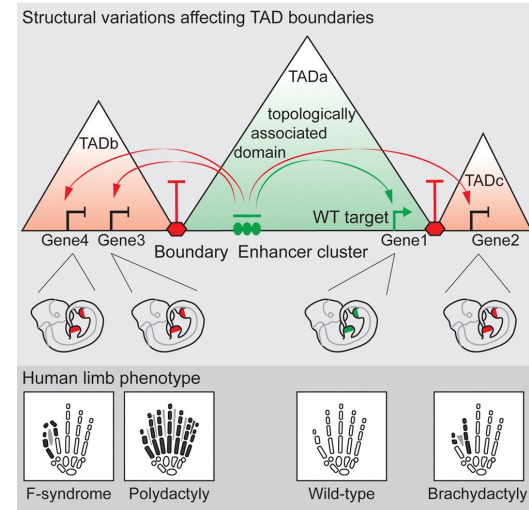
- Dense chromatin
- Gene devoid
- Methylation marks
- Toward lamina

Topological Associated Domains



High level of self-interaction:

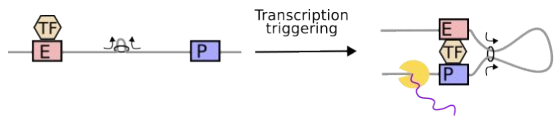
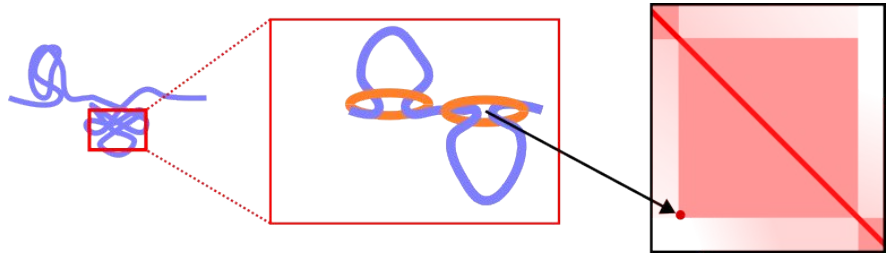
- Boundary regions insulator elements : separators
- Enriched for CTCF or architectural proteins



- Regularity units: appropriate enhancer/promoter
- Boundary disruption = inappropriate gene activation
→ Disorders (development, cancer, ...)

How do distant regulatory elements communicate with their target genes to control gene expression?

Chromatin loops: Bringing Elements Together



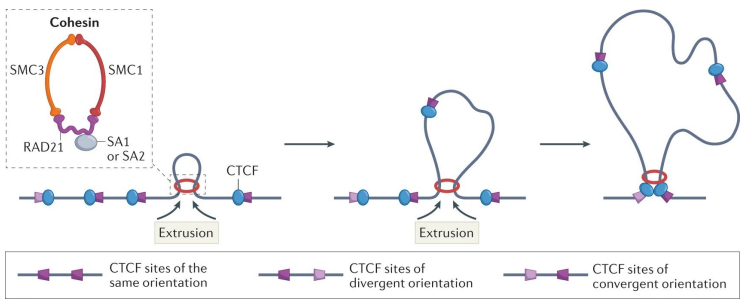
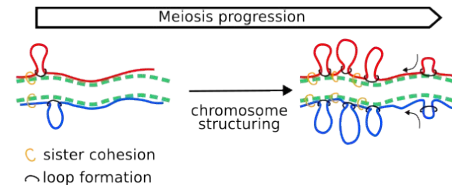
Regulatory loops

- Modulate gene expression
- Small range, gene expression precise control
- Dynamic: TF, chromatine remodelers, signaling

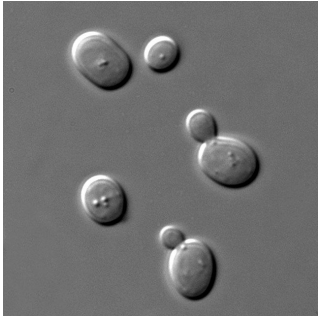
Dual role, different scales but shared molecular mechanisms

Structural loops

- Establish, maintain territories
- Long range, overall genome structuration
- More stable → slower timescale (cellular cycle)



S. cerevisiae as a model for 3D genomics studies



- ~ 12 millions bp
- ~ 6000 genes
- Haploid and diploid states
- Easy to handle

